

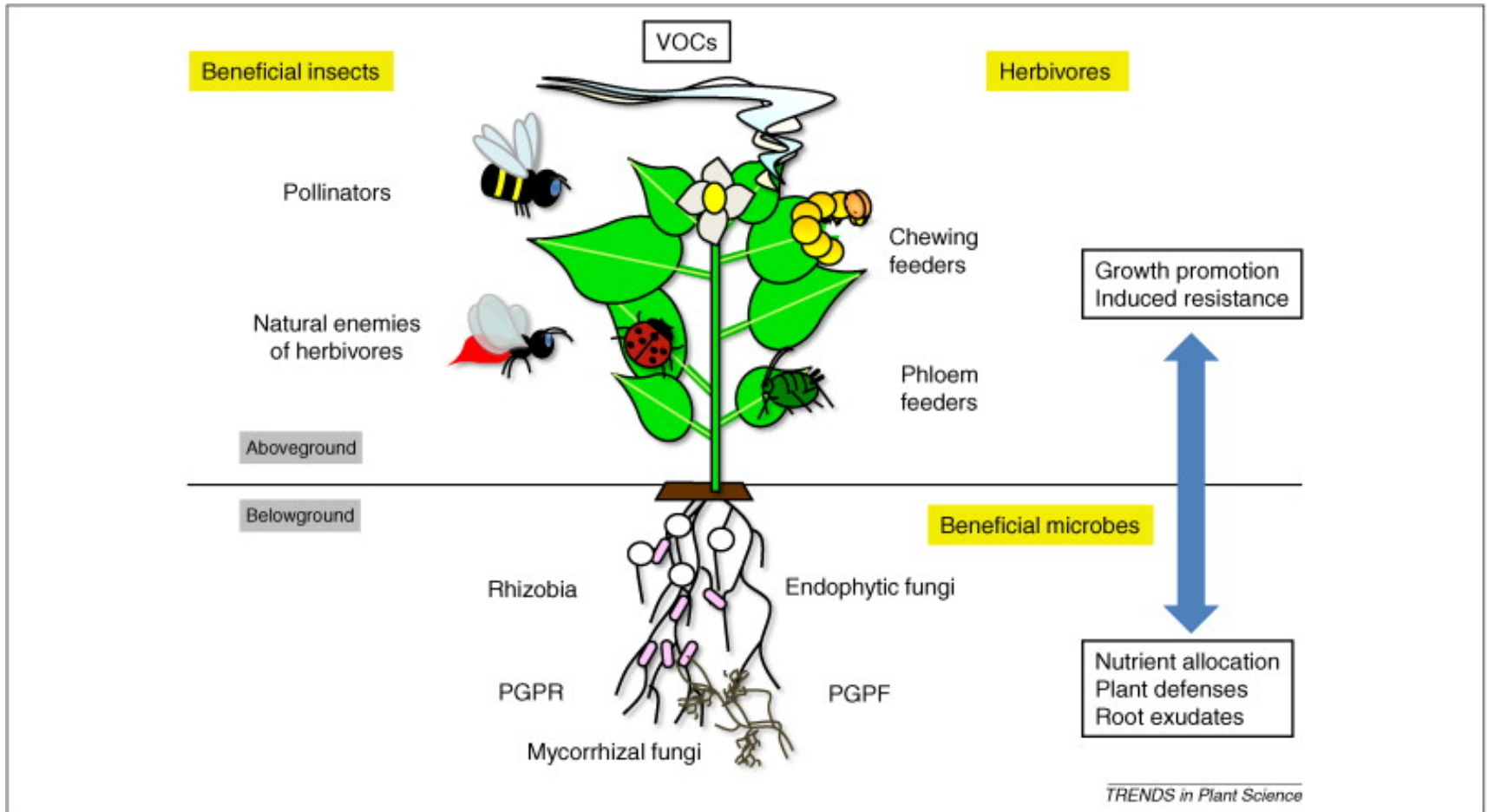
Immunity, stress and sub-lethal effects of neonicotinoids

Francesco Pennacchio

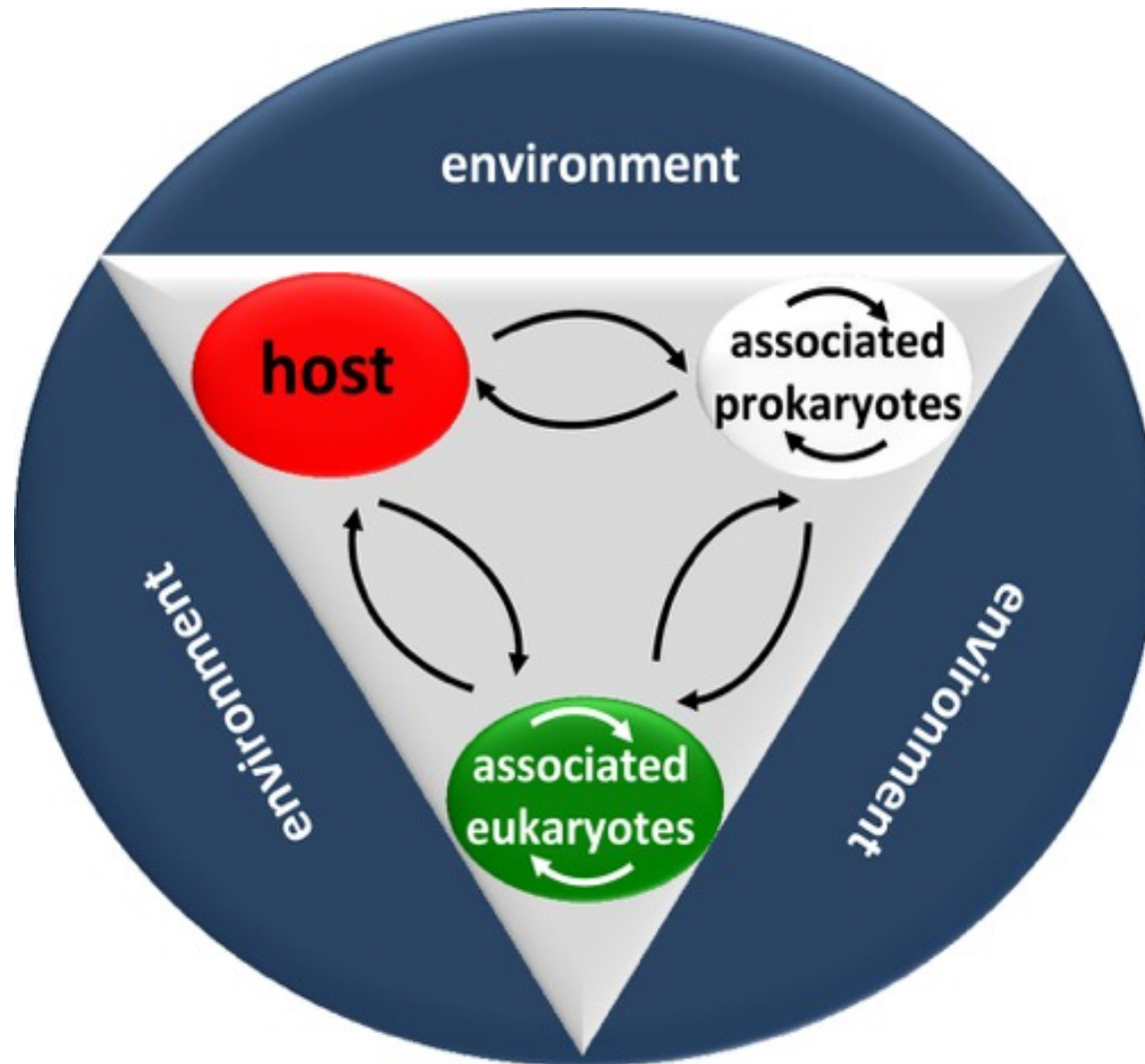
*University of Napoli “Federico II”
Department of Agricultural Sciences
Portici (NA) - ITALY*



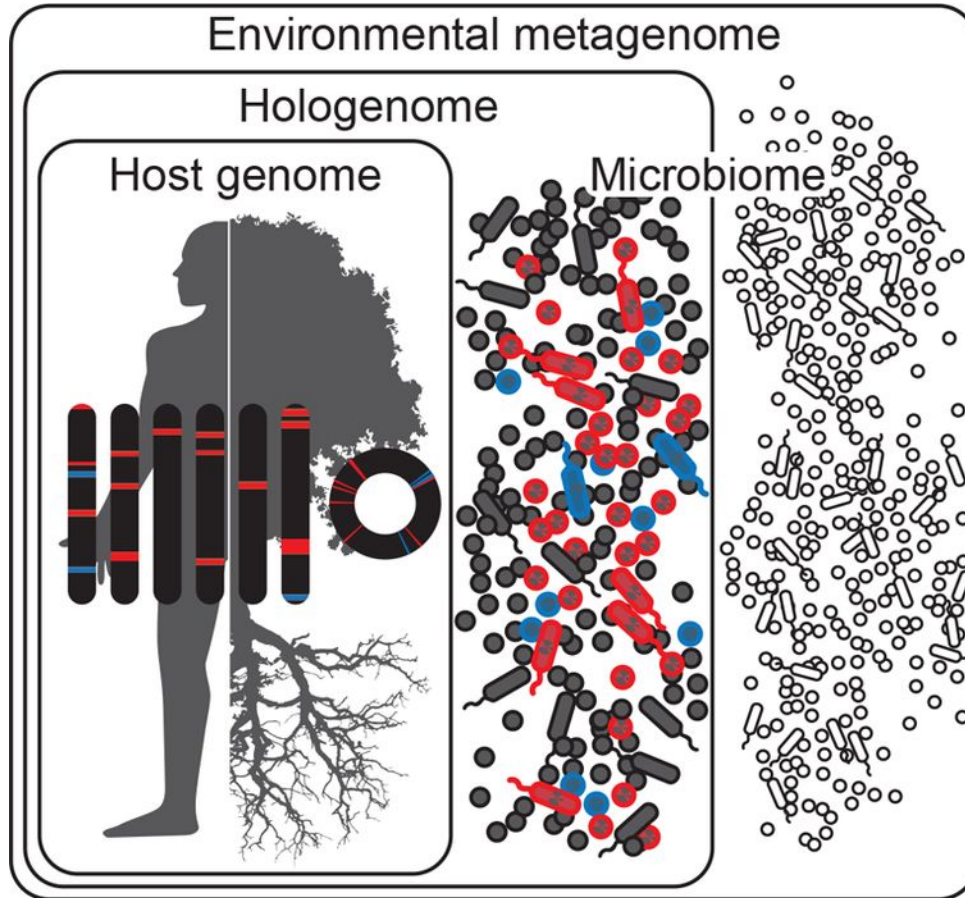
Multitrophic Interactions





Interactions at metaorganism level





Holobiont phenotype is controlled by the hologenome



 Host and symbiont genes that alone and/or together affect a holobiont phenotype

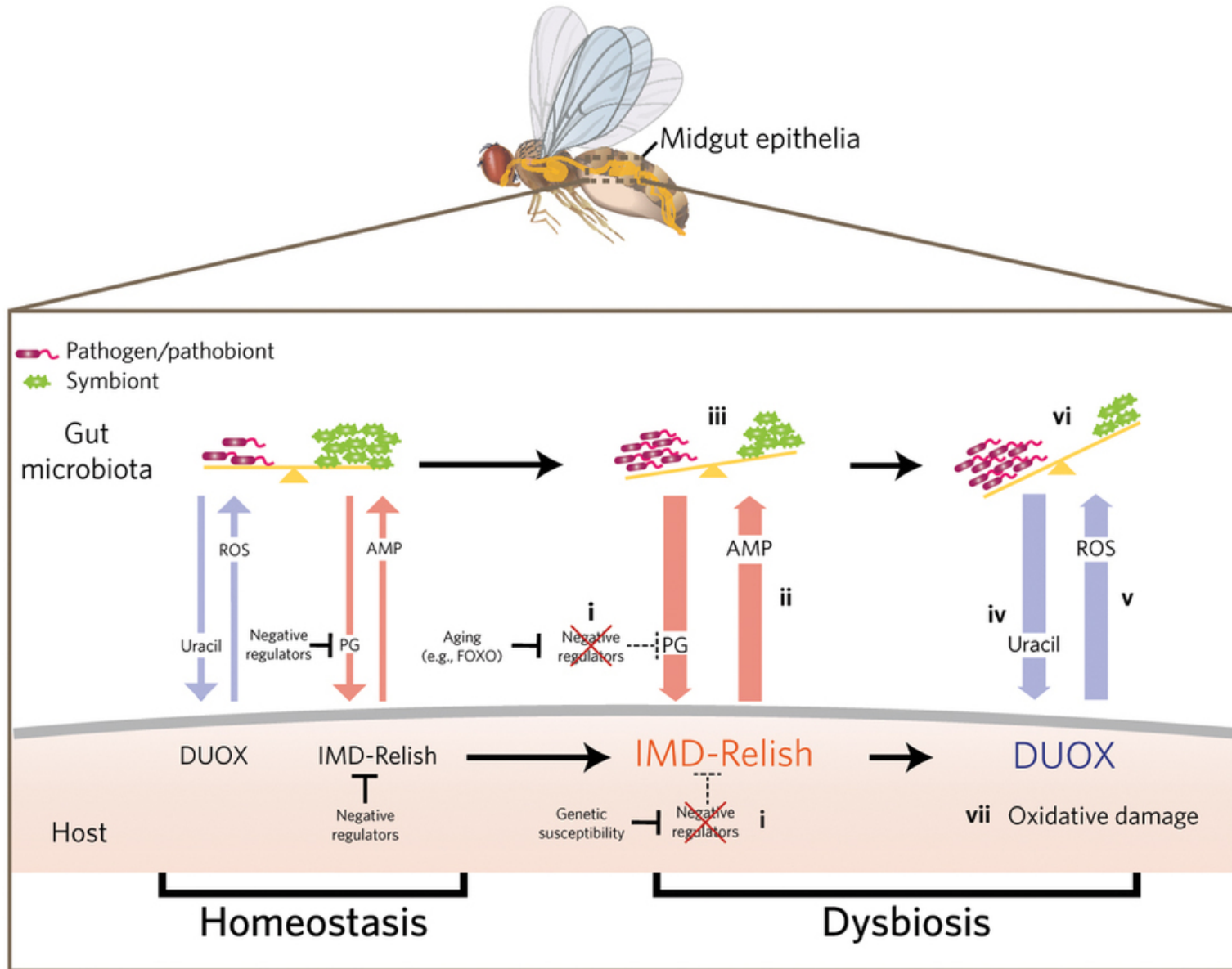
 Coevolved host and symbiont genes that affect a holobiont phenotype

 Host genes and symbionts that do not affect a holobiont phenotype

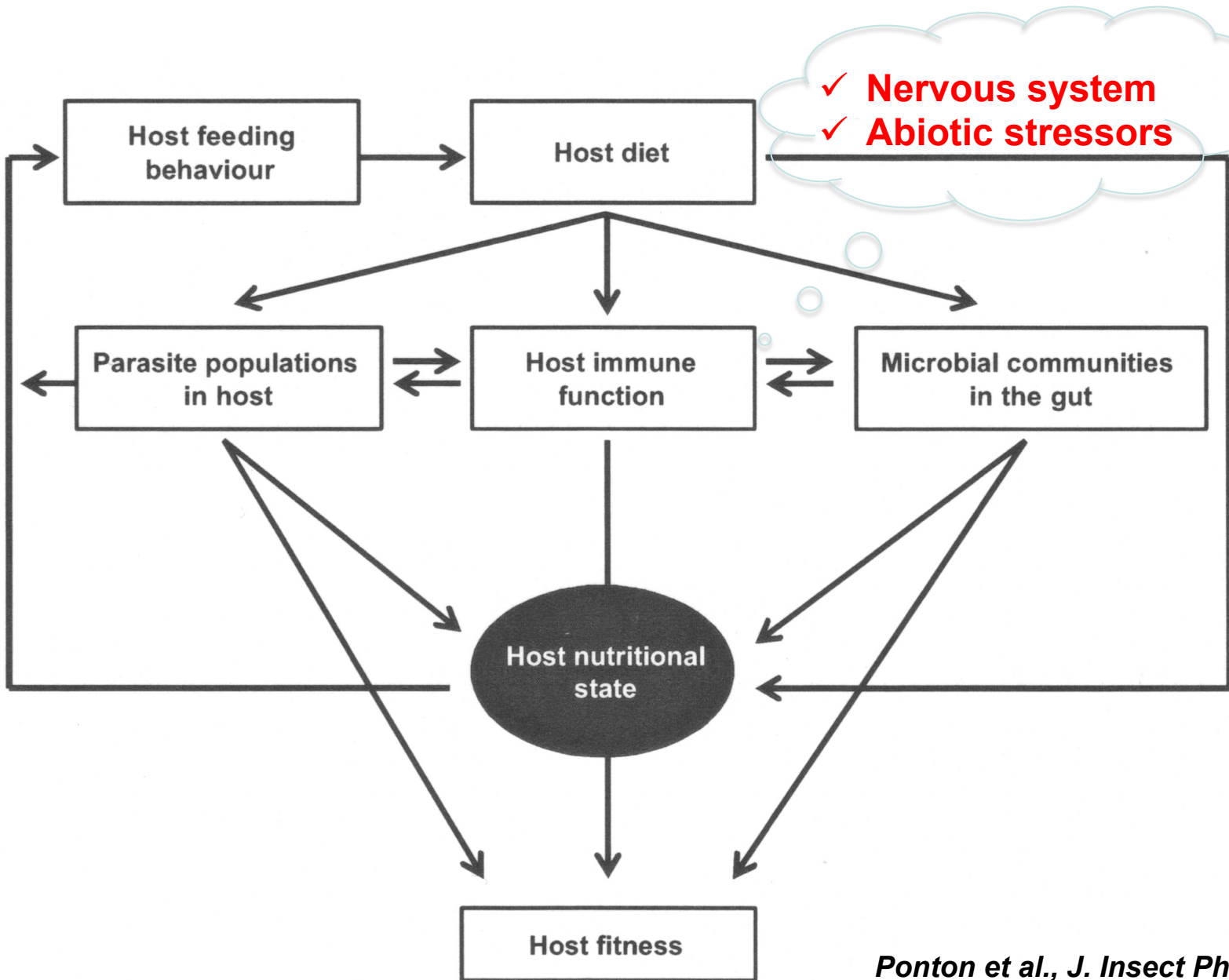
 Environmental microbes that are not part of the holobiont

Kevin R. Theis et al. *mSystems* 2016;1:e00028-16

Gut Microbiota and Immunity



Multifactorial regulation of immunity

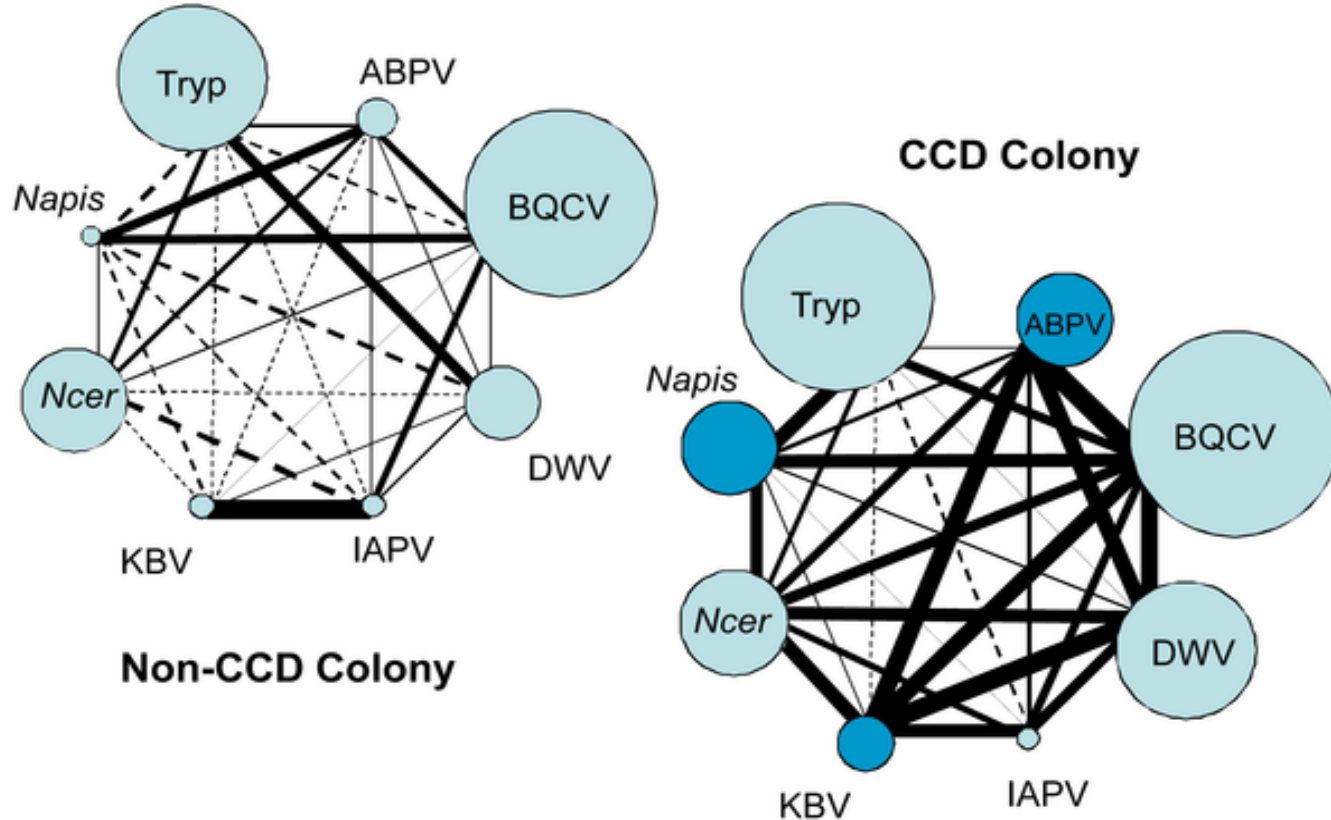


Stress and Honeybee Immunity



EU – FP7

Pathogen loads are highly covariant in collapsing colonies



- ✓ Increased susceptibility to a diverse set of pathogens
- ✓ Co-infections can act synergistically

>20 km

The induced collapse experiment



1.6 km



Same environment
but no interactions

Observations on:

- colony strength
- mite infestation
- bee mortality
- pathogens



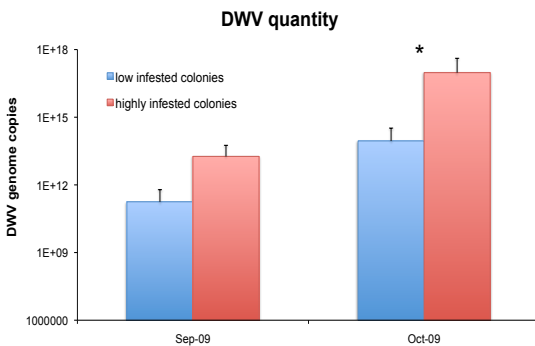
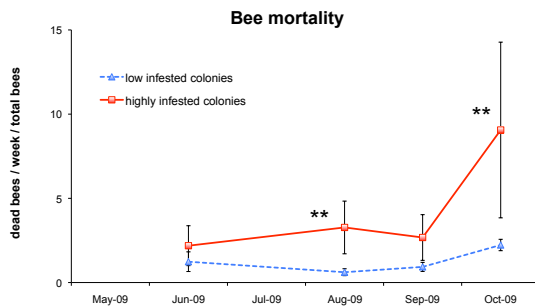
low infested colonies (LIC)



highly infested colonies (HIC)

Expression of immune genes

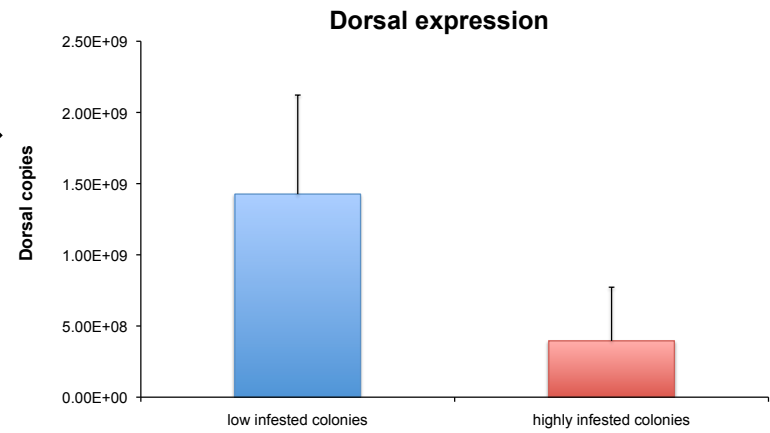
Bees from HIC



RNA-seq

Gene Name	Gene Family/Pathway	HIC1	HIC2	HIC3	HIC4	HIC5
AbpA		0.92	0.92	0.92	0.92	0.92
AbpA2		0.92	0.92	0.92	0.92	0.92
AbpA3		0.92	0.92	0.92	0.92	0.92
AbpA4		0.92	0.92	0.92	0.92	0.92
AbpA5		0.92	0.92	0.92	0.92	0.92
AbpA6		0.92	0.92	0.92	0.92	0.92
AbpA7		0.92	0.92	0.92	0.92	0.92
AbpA8		0.92	0.92	0.92	0.92	0.92
AbpA9		0.92	0.92	0.92	0.92	0.92
AbpA10		0.92	0.92	0.92	0.92	0.92
AbpA11		0.92	0.92	0.92	0.92	0.92
AbpA12		0.92	0.92	0.92	0.92	0.92
AbpA13		0.92	0.92	0.92	0.92	0.92
AbpA14		0.92	0.92	0.92	0.92	0.92
AbpA15		0.92	0.92	0.92	0.92	0.92
AbpA16		0.92	0.92	0.92	0.92	0.92
AbpA17		0.92	0.92	0.92	0.92	0.92
AbpA18		0.92	0.92	0.92	0.92	0.92
AbpA19		0.92	0.92	0.92	0.92	0.92
AbpA20		0.92	0.92	0.92	0.92	0.92
AbpA21		0.92	0.92	0.92	0.92	0.92
AbpA22		0.92	0.92	0.92	0.92	0.92
AbpA23		0.92	0.92	0.92	0.92	0.92
AbpA24		0.92	0.92	0.92	0.92	0.92
AbpA25		0.92	0.92	0.92	0.92	0.92
AbpA26		0.92	0.92	0.92	0.92	0.92
AbpA27		0.92	0.92	0.92	0.92	0.92
AbpA28		0.92	0.92	0.92	0.92	0.92
AbpA29		0.92	0.92	0.92	0.92	0.92
AbpA30		0.92	0.92	0.92	0.92	0.92
AbpA31		0.92	0.92	0.92	0.92	0.92
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AbpA34		0.92	0.92	0.92	0.92	0.92
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AbpA36		0.92	0.92	0.92	0.92	0.92
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AbpA72		0.92	0.92	0.92	0.92	0.92
AbpA73		0.92	0.92	0.92	0.92	0.92
AbpA74		0.92	0.92	0.92	0.92	0.92
AbpA75		0.92	0.92	0.92	0.92	0.92
AbpA76		0.92	0.92	0.92	0.92	0.92
AbpA77		0.92	0.92	0.92	0.92	0.92
AbpA78		0.92	0.92	0.92	0.92	0.92
AbpA79		0.92	0.92	0.92	0.92	0.92
AbpA80		0.92	0.92	0.92	0.92	0.92
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AbpA82		0.92	0.92	0.92	0.92	0.92
AbpA83		0.92	0.92	0.92	0.92	0.92
AbpA84		0.92	0.92	0.92	0.92	0.92
AbpA85		0.92	0.92	0.92	0.92	0.92
AbpA86		0.92	0.92	0.92	0.92	0.92
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AbpA89		0.92	0.92	0.92	0.92	0.92
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AbpA91		0.92	0.92	0.92	0.92	0.92
AbpA92		0.92	0.92	0.92	0.92	0.92
AbpA93		0.92	0.92	0.92	0.92	0.92
AbpA94		0.92	0.92	0.92	0.92	0.92
AbpA95		0.92	0.92	0.92	0.92	0.92
AbpA96		0.92	0.92	0.92	0.92	0.92
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AbpA98		0.92	0.92	0.92	0.92	0.92
AbpA99		0.92	0.92	0.92	0.92	0.92
AbpA100		0.92	0.92	0.92	0.92	0.92

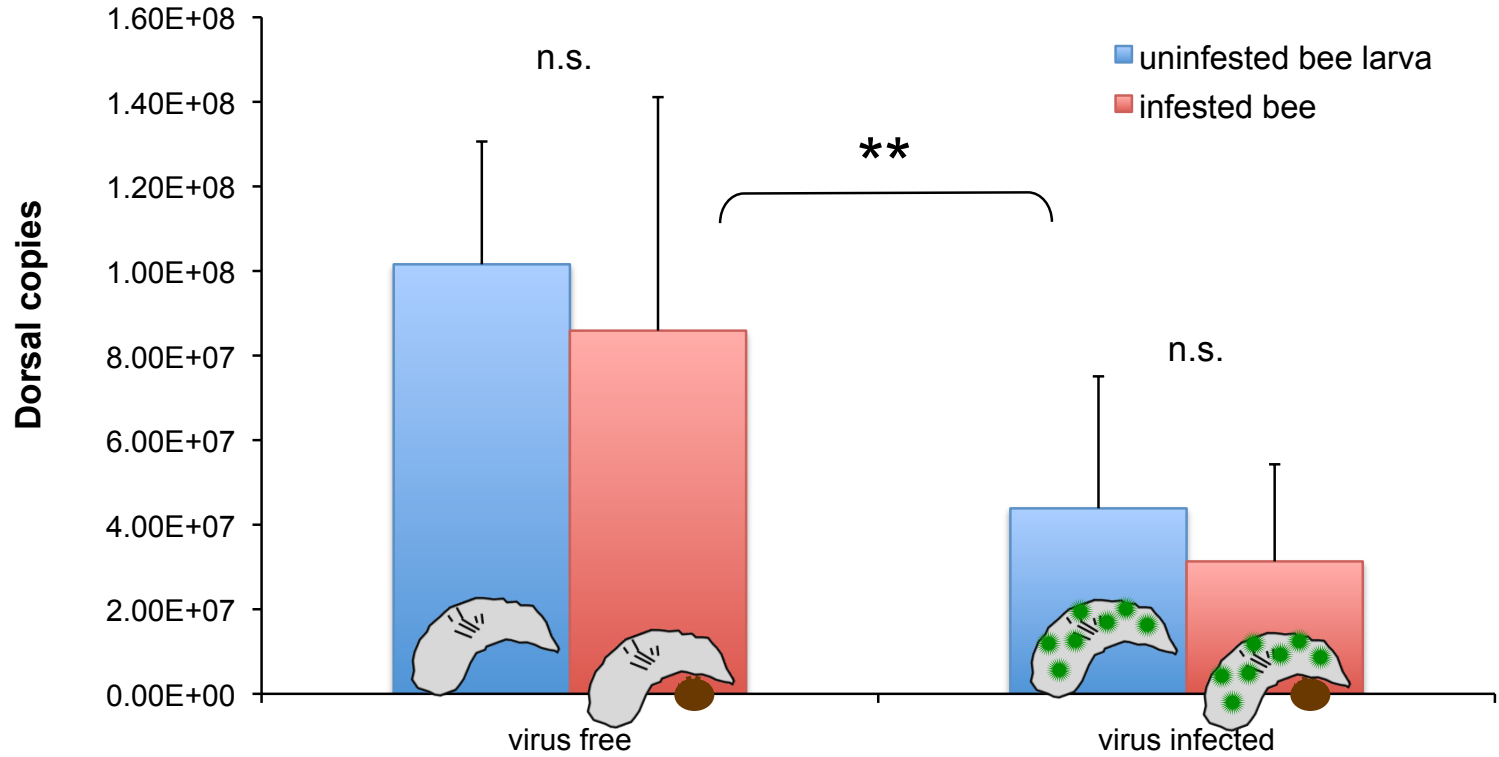
Real time RT-PCR of selected genes



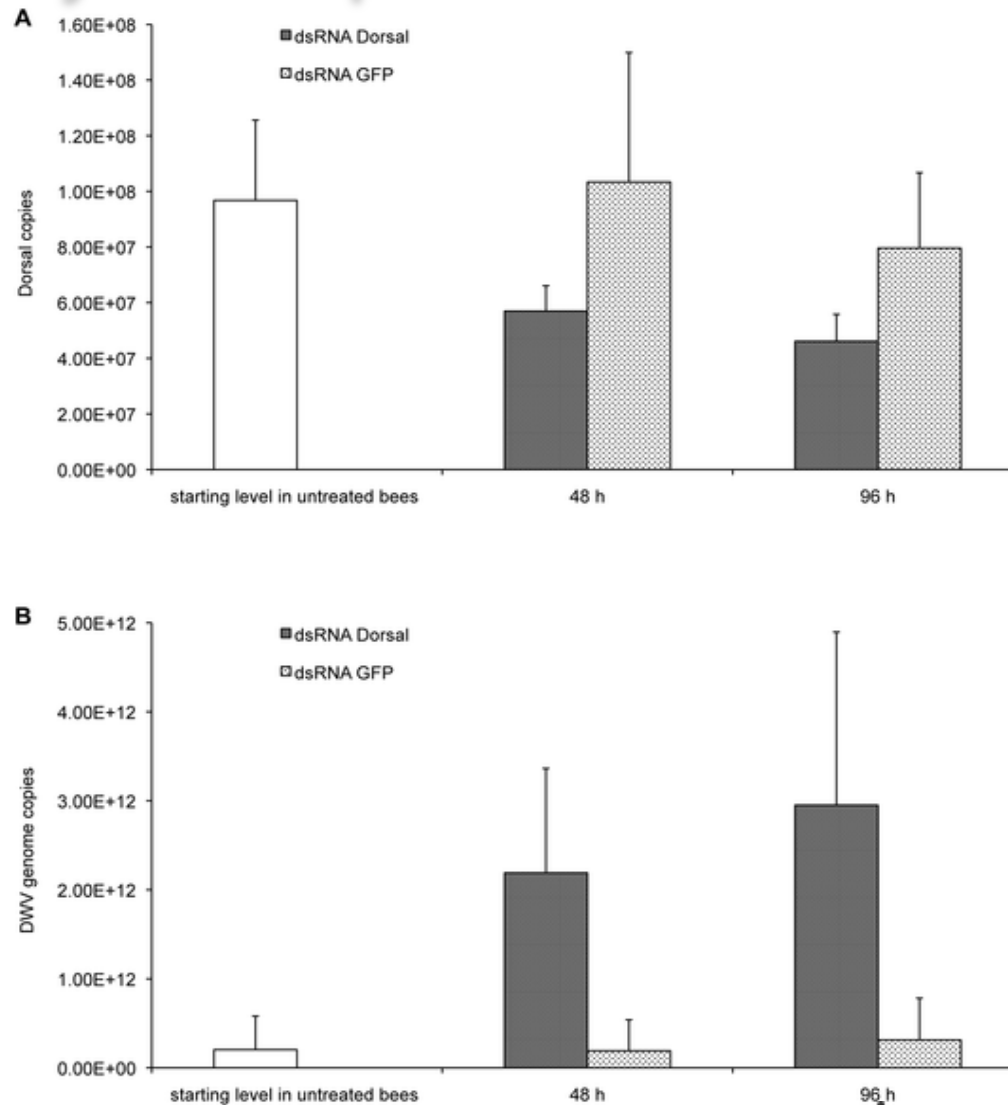
Transcription of *dorsal-1A* (*NF-kB*) is highly down-regulated in HIC

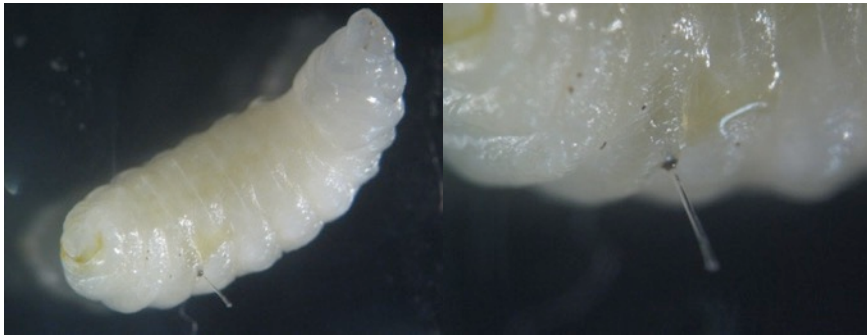
Dorsal expression is influenced by DWV

Influence on Dorsal: *Varroa* or DWV?



The down-regulation of the transcription factor dorsal-1A by RNAi promotes DWV replication





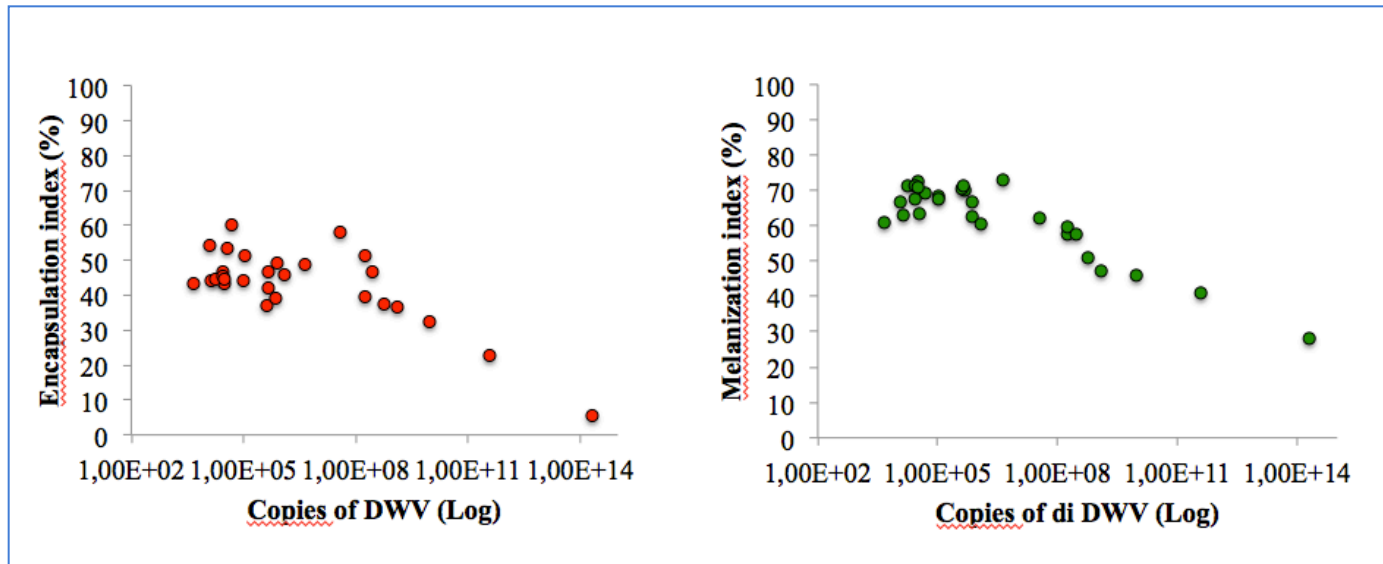
Nylon thread implantation

DWV titer positively correlates with honeybee immunosuppression

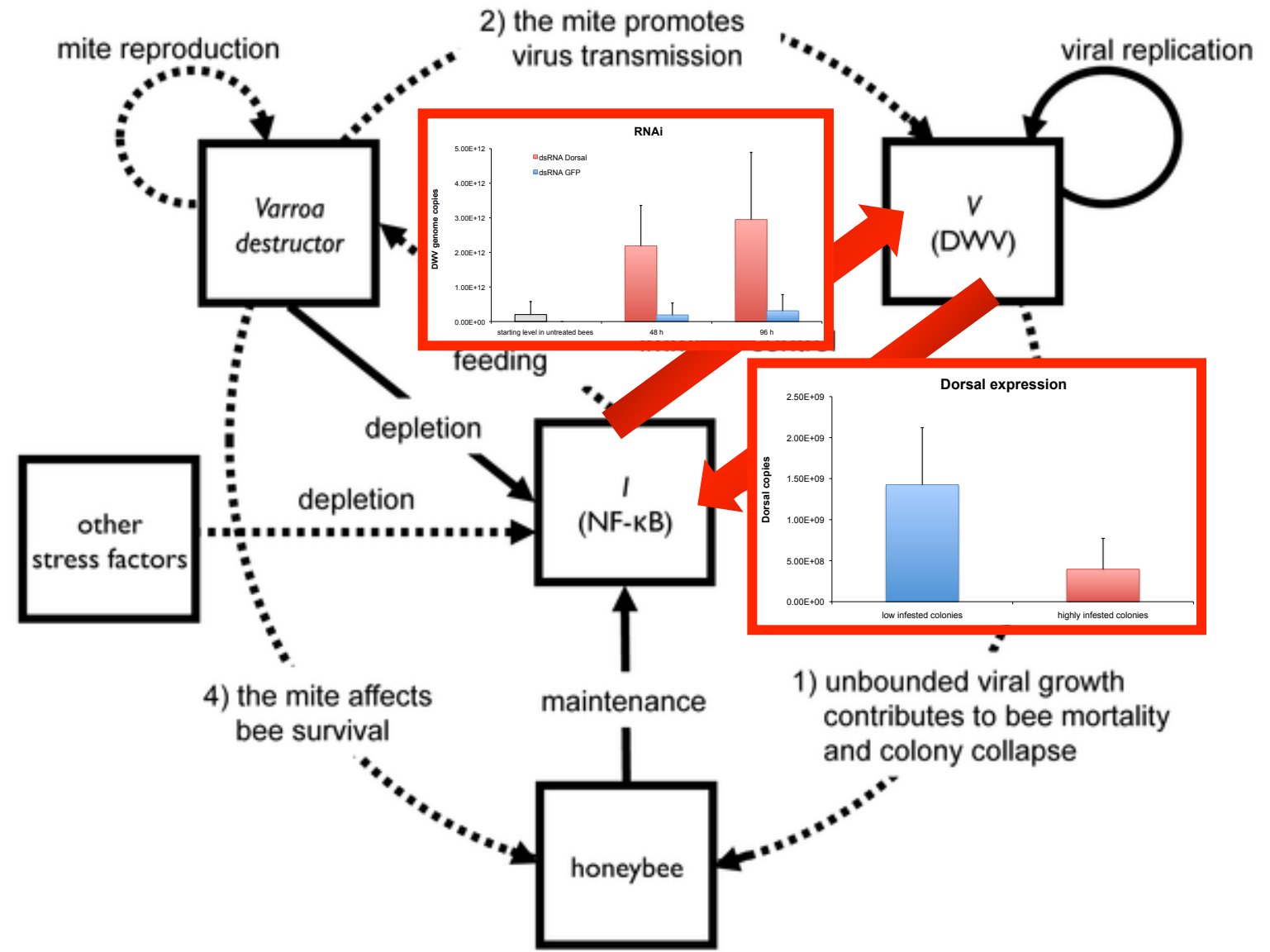


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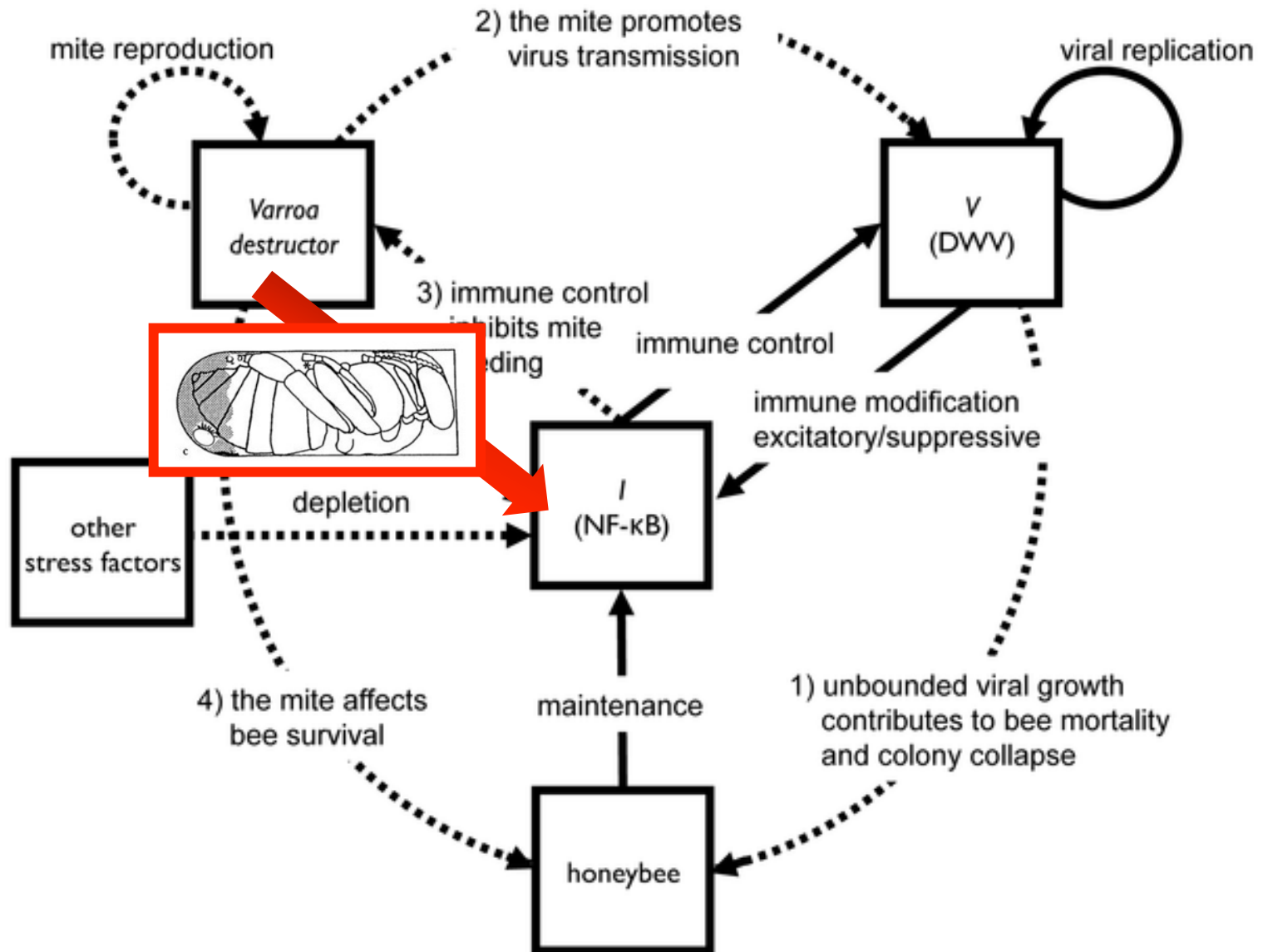
+ DWV



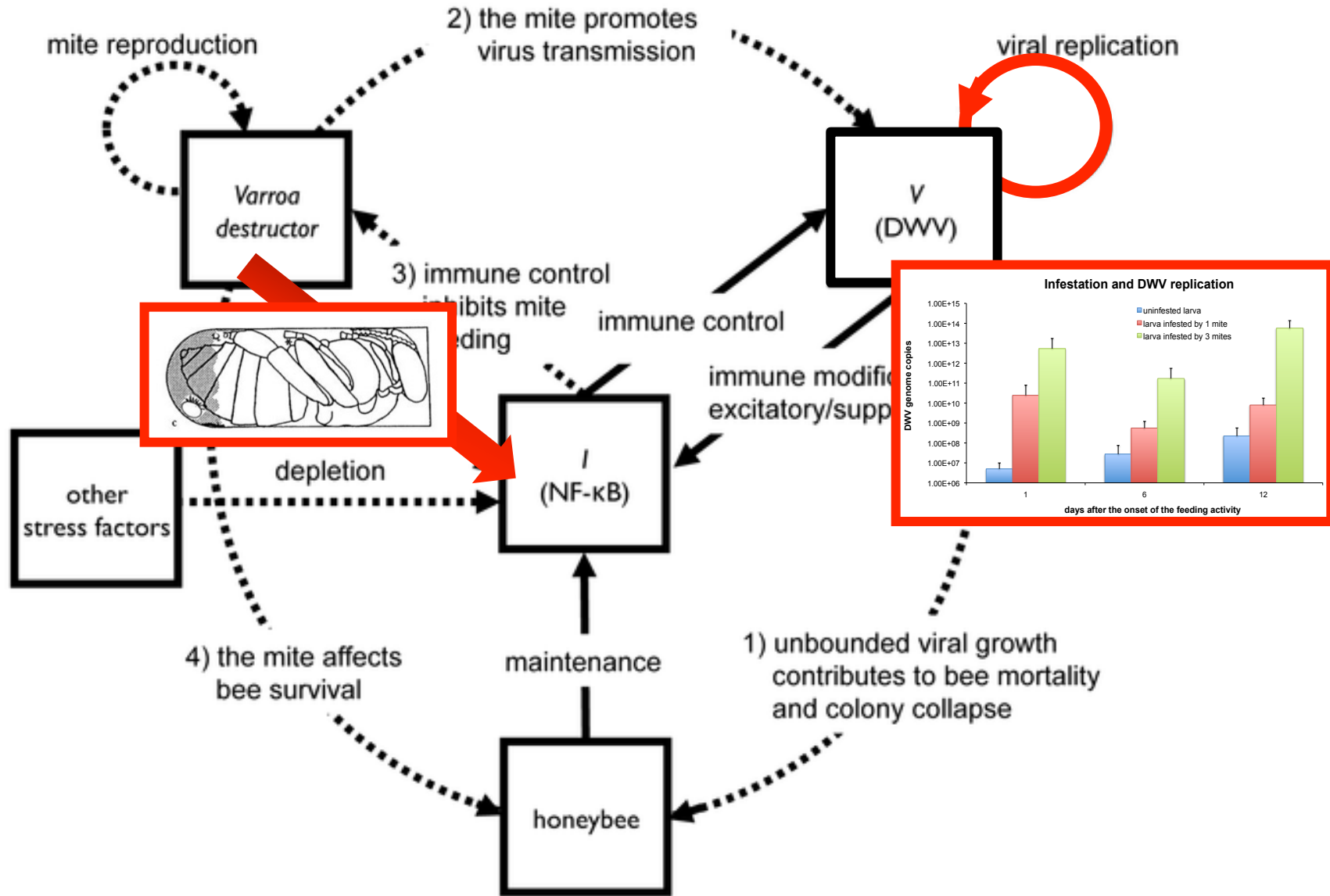
A delicate balance underpins covert DWV infections



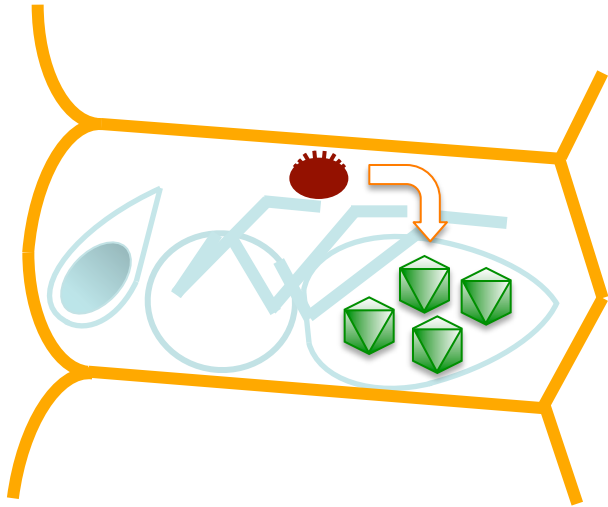
Varroa triggers immune reactions associated with a severe metabolic stress...



...which promote viral replication

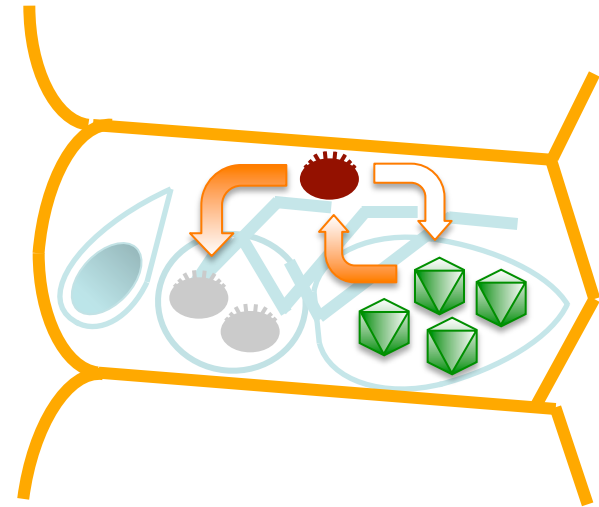


Varroa acts as a vector
and promotes DWV
replication



+

Bee immunosuppression by DWV
favours Varroa feeding and enhances
mite's fitness



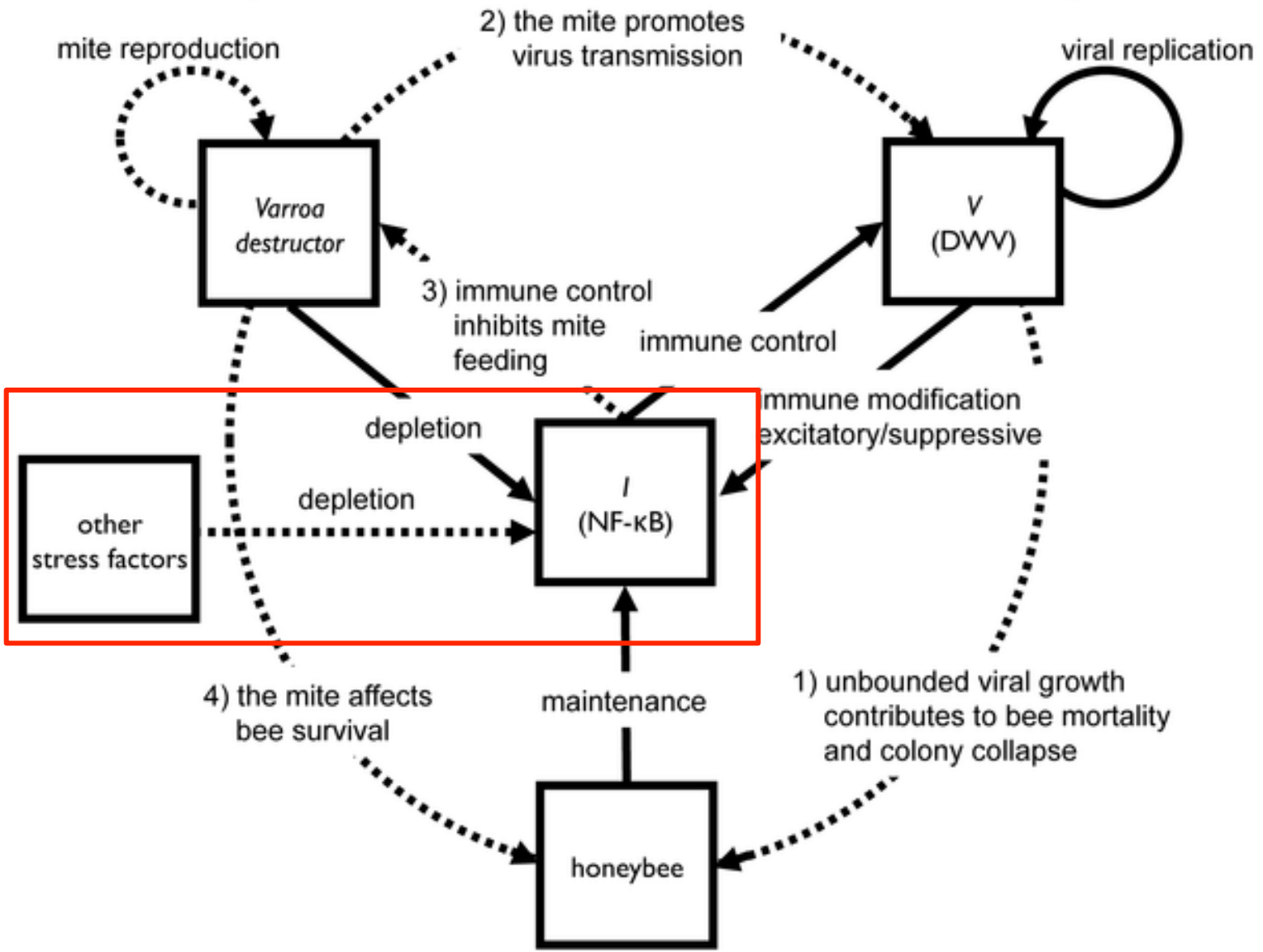
A mutualistic symbiosis between a parasitic mite and a pathogenic virus undermines honey bee immunity and health

Gennaro Di Prisco^{a,1}, Desiderato Annoscia^{b,1}, Marina Margiotta^a, Rosalba Ferrara^a, Paola Varricchio^a, Virginia Zanni^b, Emilio Caprio^a, Francesco Nazzi^{b,2}, and Francesco Pennacchio^{a,2}

^aDipartimento di Agraria, Laboratorio di Entomologia "E. Tremblay," Università degli Studi di Napoli "Federico II," 80055 Portici (NA), Italy;

and ^bDipartimento di Scienze AgroAlimentari Ambientali e Animali, Università degli Studi di Udine, 33100 Udine, Italy

Do stress factors acting on bee immunocompetence influence DWV replication?





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Transcriptomic and proteomic effects of a neonicotinoid insecticide mixture in the marine mussel (*Mytilus galloprovincialis*, Lam.)

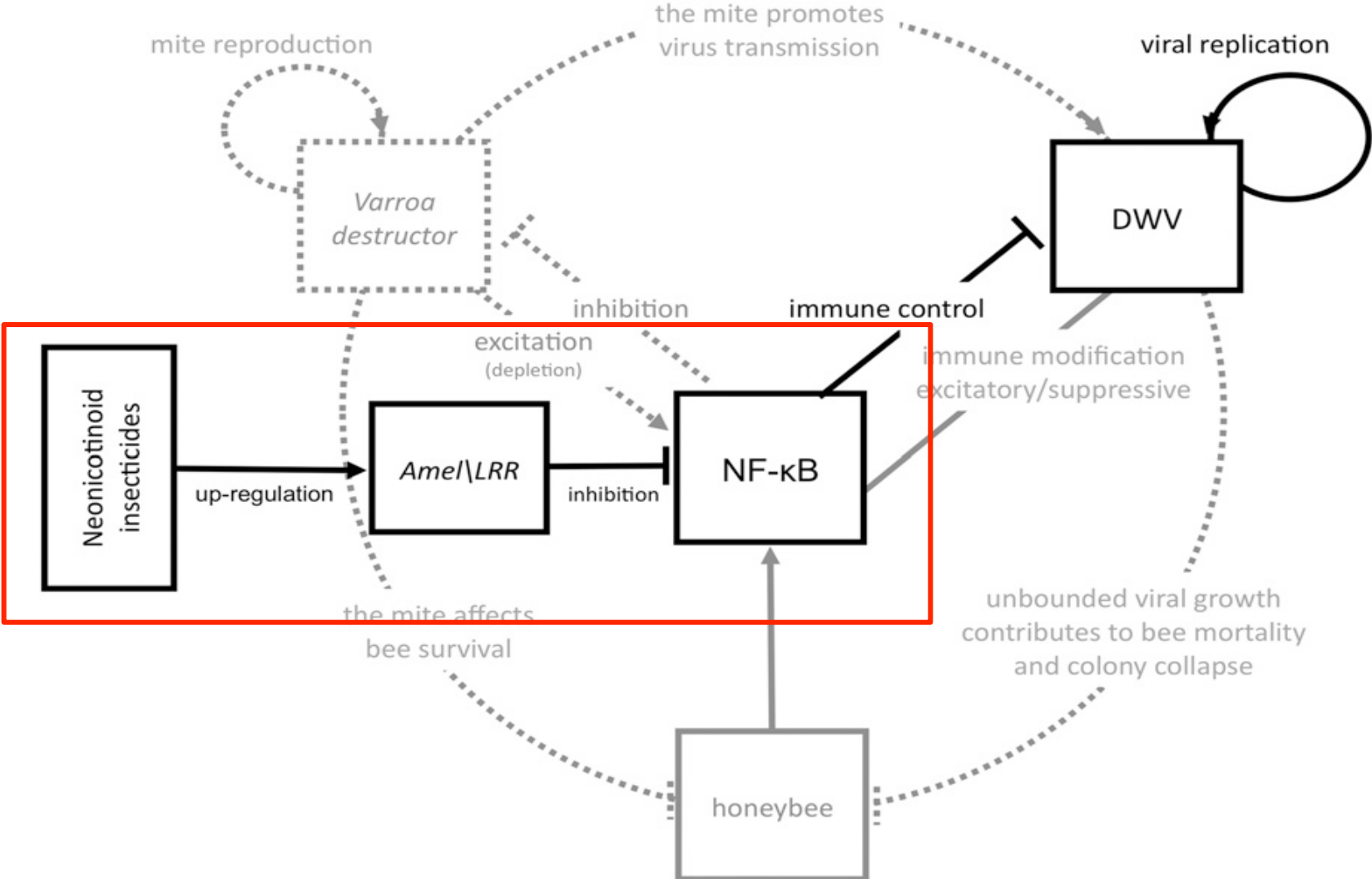
Francesco Dondero ^{a,*}, Alessandro Negri ^{a,1}, Lara Boatti ^a, Francesco Marsano ^a, Flavio Mignone ^b, Aldo Viarengo ^a

^a Department of Environmental and Life Sciences, Università del Piemonte Orientale Amedeo Avogadro, Alessandria, Italy

^b Department of Structural Chemistry, University of Milan, Milan, Italy

“Thiacloprid elicited the modulation of gene transcription and mRNA metabolic processes: three ribonucleoproteins, and two transcription factors (mflj00348 protein, also known as “caterpillar” in mammals) were identified. The latter sequences, in human cells, may modulate T-cell activation, decrease the transcription of genes that are normally up-regulated after T-cell stimulation and delays degradation of NFKBIA/IKBA”

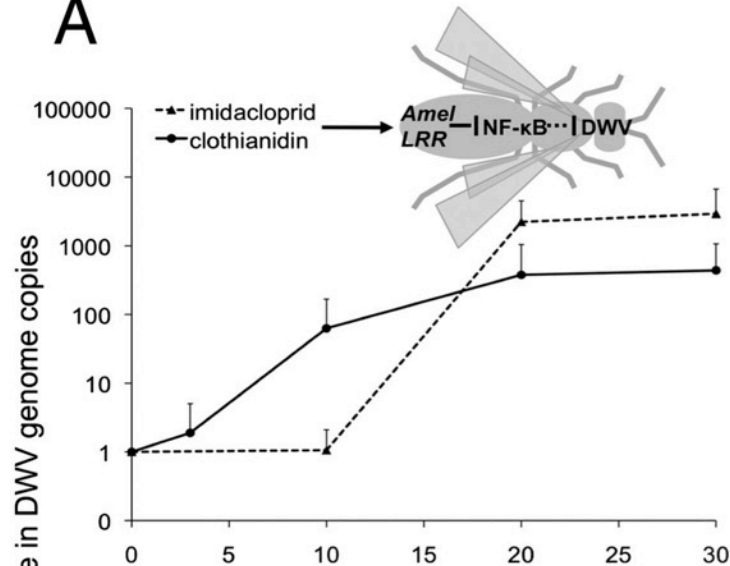
Neonicotinoid impact on honey bee immunity



Effect of insecticides on DWV replication in honey bees bearing covert infections

Topical

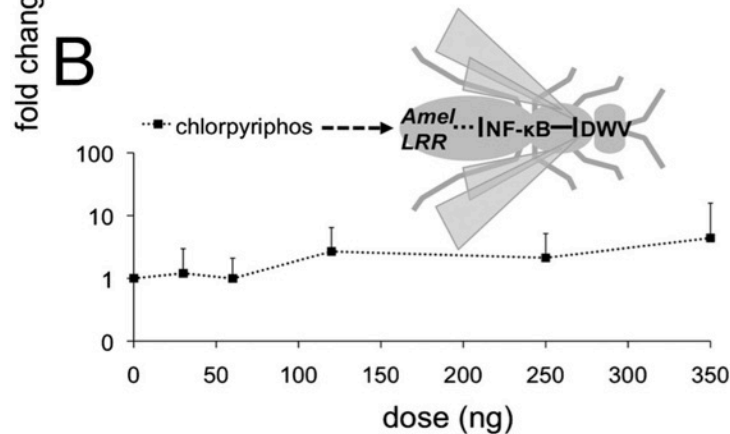
A



insecticide treatment

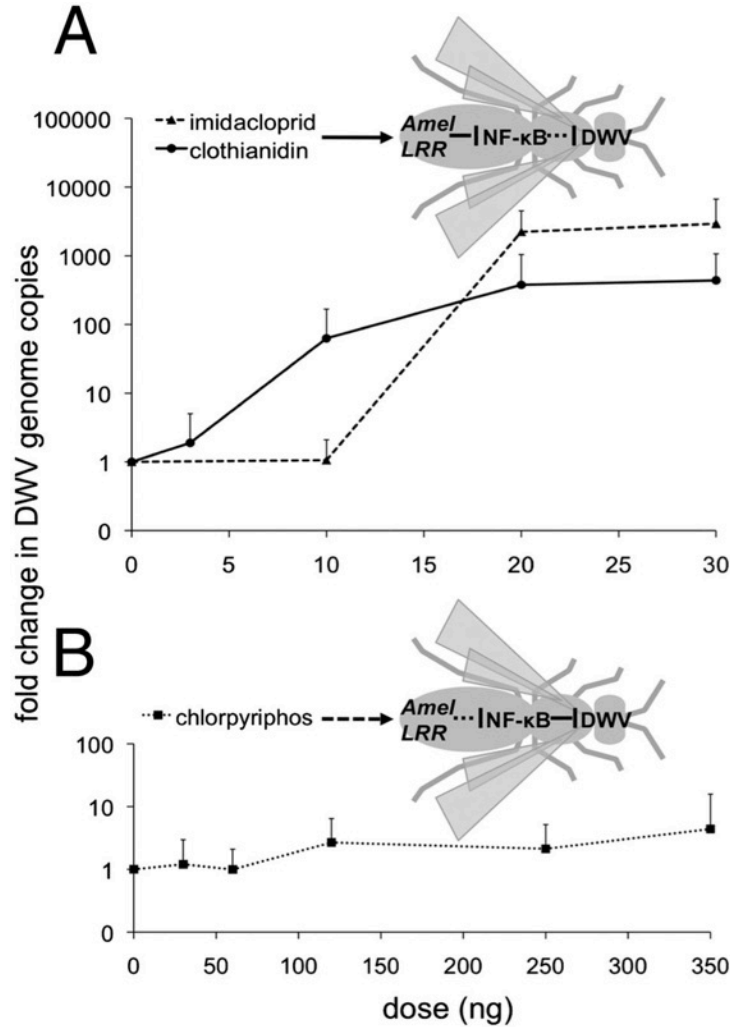


B

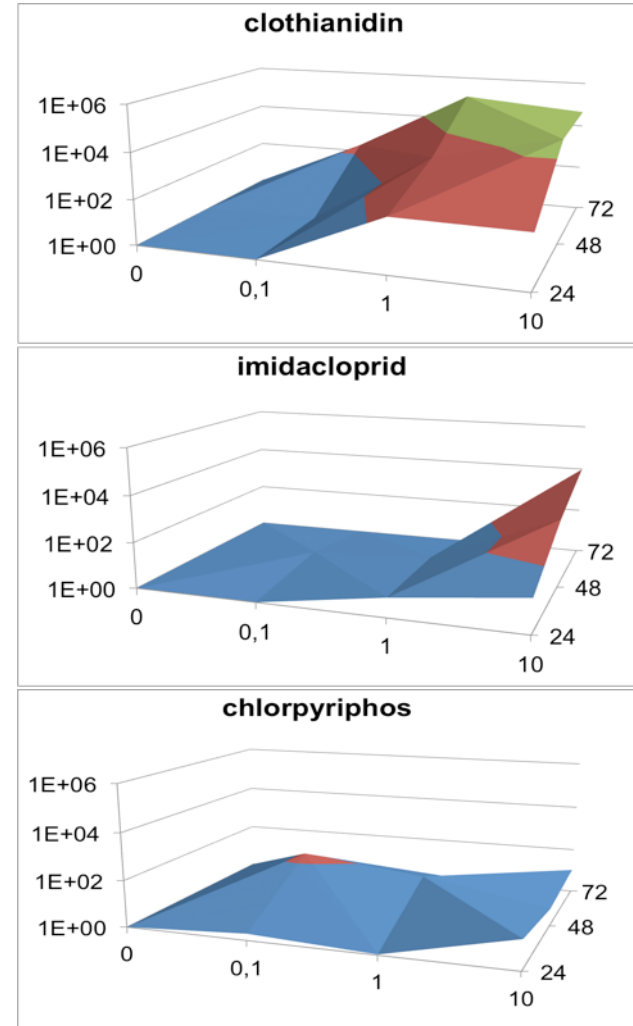


Effect of insecticides on DWV replication in honey bees bearing covert infections

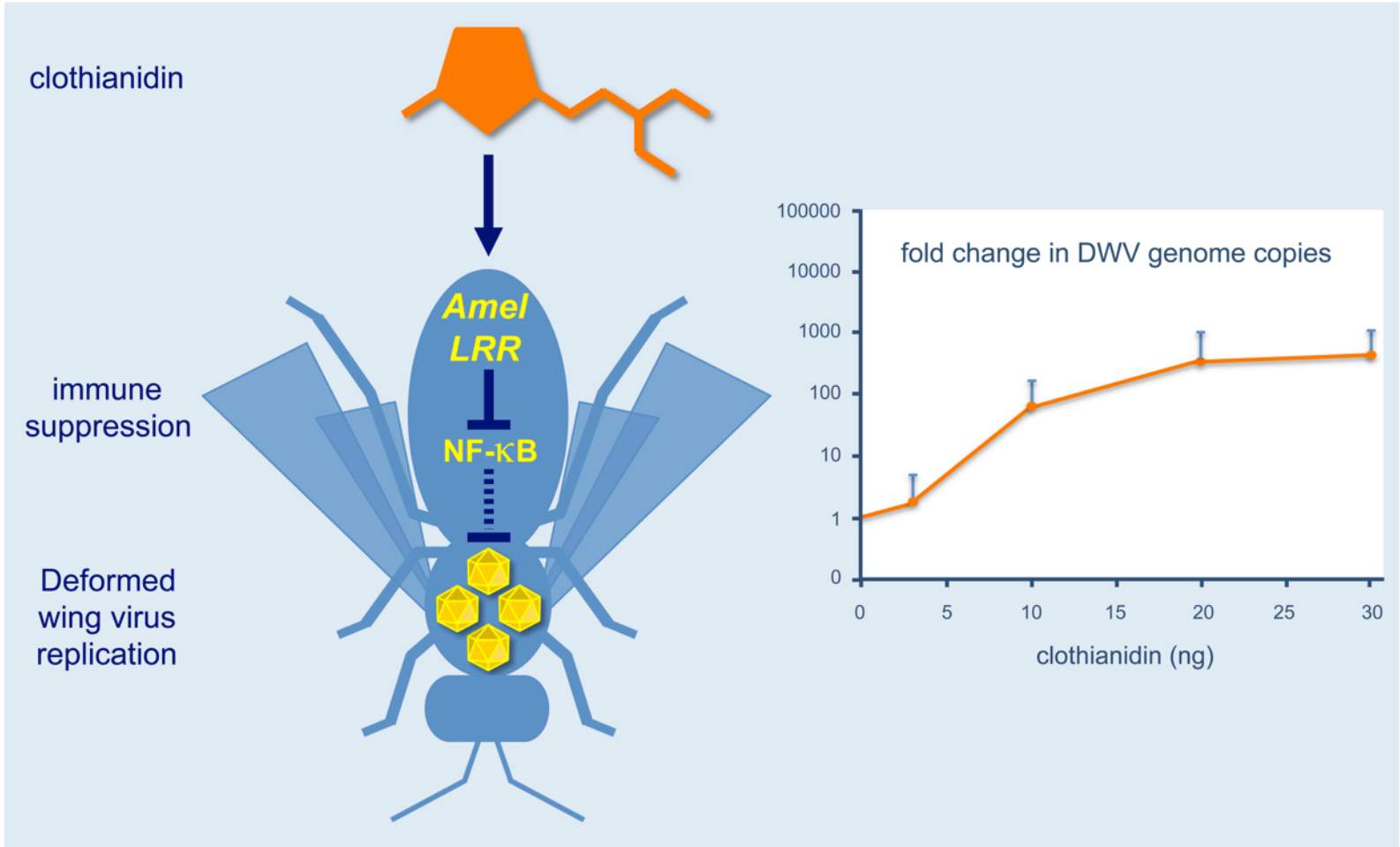
Topical



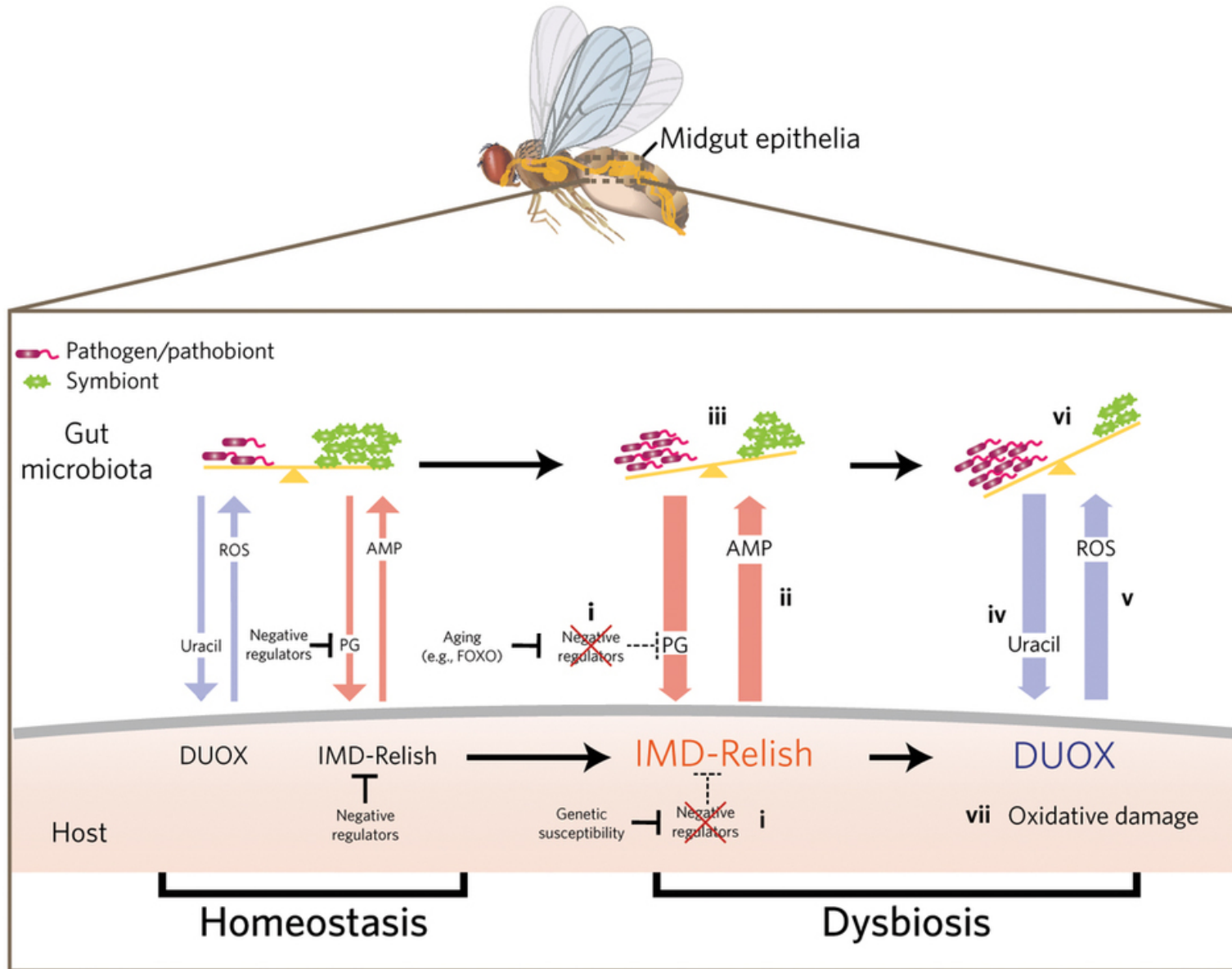
Oral



Neonicotinoids and honey bee antiviral immunity



Gut Microbiota and Immunity




SCIENTIFIC REPORTS



OPEN

Neonicotinoid-induced pathogen susceptibility is mitigated by *Lactobacillus plantarum* immune stimulation in a *Drosophila melanogaster* model

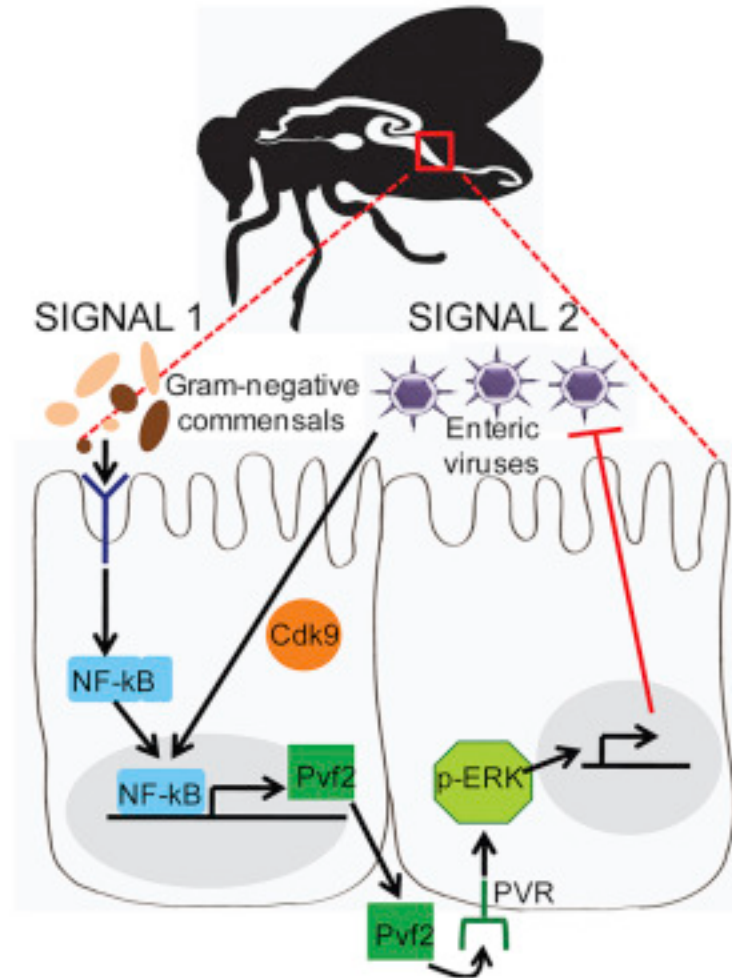
Brendan A. Daisley^{1,2}, Mark Trinder ^{1,2}, Tim W. McDowell³, Hylke Welle^{1,2,4}, Josh S. Dube², Sohrab N. Ali^{2,5}, Hon S. Leong^{2,6}, Mark W. Sumarah³ & Gregor Reid ^{1,2,6}

Received: 19 December 2016

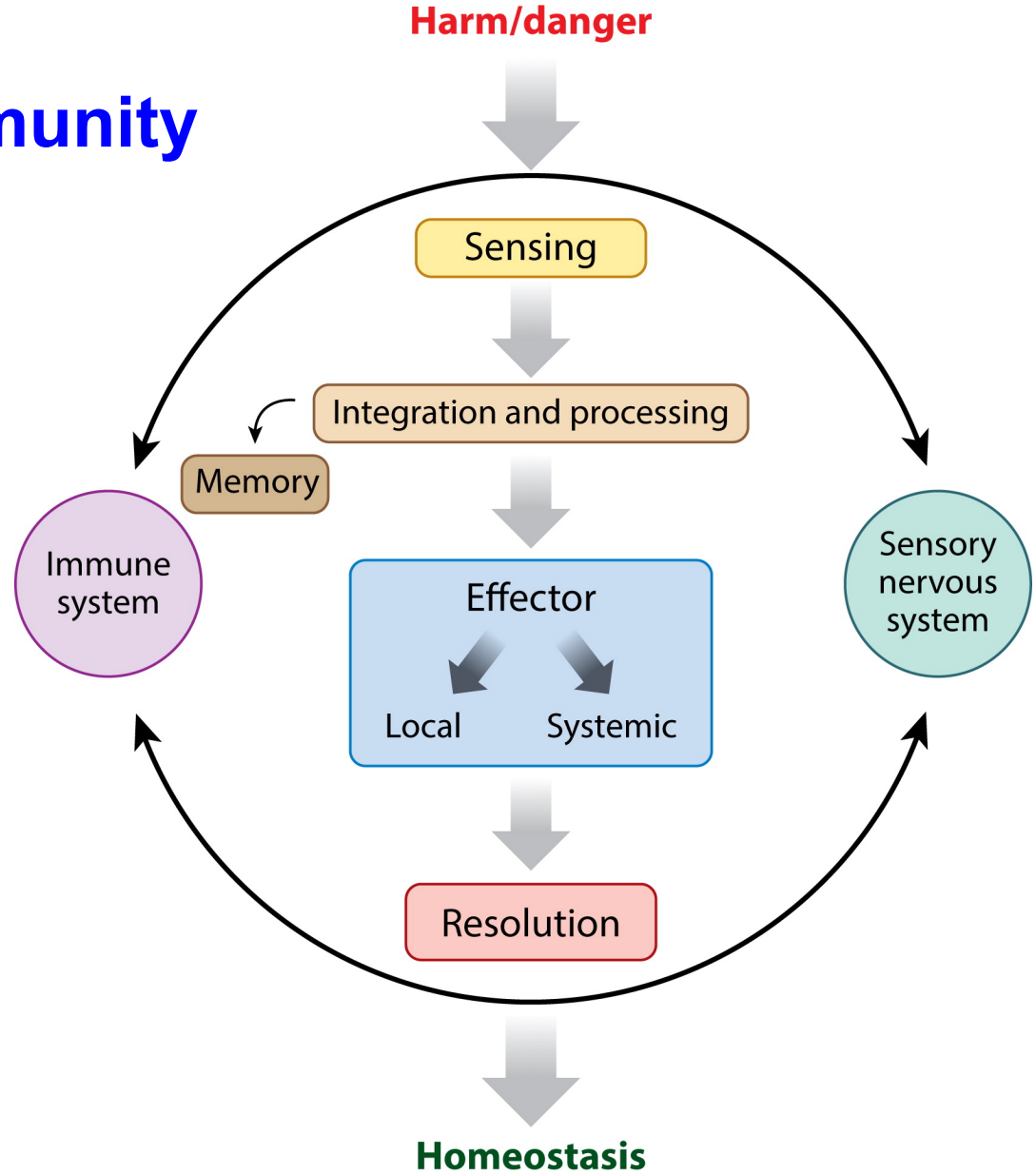
Accepted: 19 April 2017

Published online: 02 June 2017

Microbiota priming of antiviral immunity



Neural control of immunity



Talbot S, et al. 2016.

Annu. Rev. Immunol. 34:421–47

SCIENTIFIC REPORTS



OPEN

The neonicotinoid insecticide Clothianidin adversely affects immune signaling in a human cell line

Gennaro Di Prisco, Marco Iannaccone, Flora Ianniello, Rosalba Ferrara, Emilio Caprio , Francesco Pennacchio & Rosanna Capparelli

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Life Sciences

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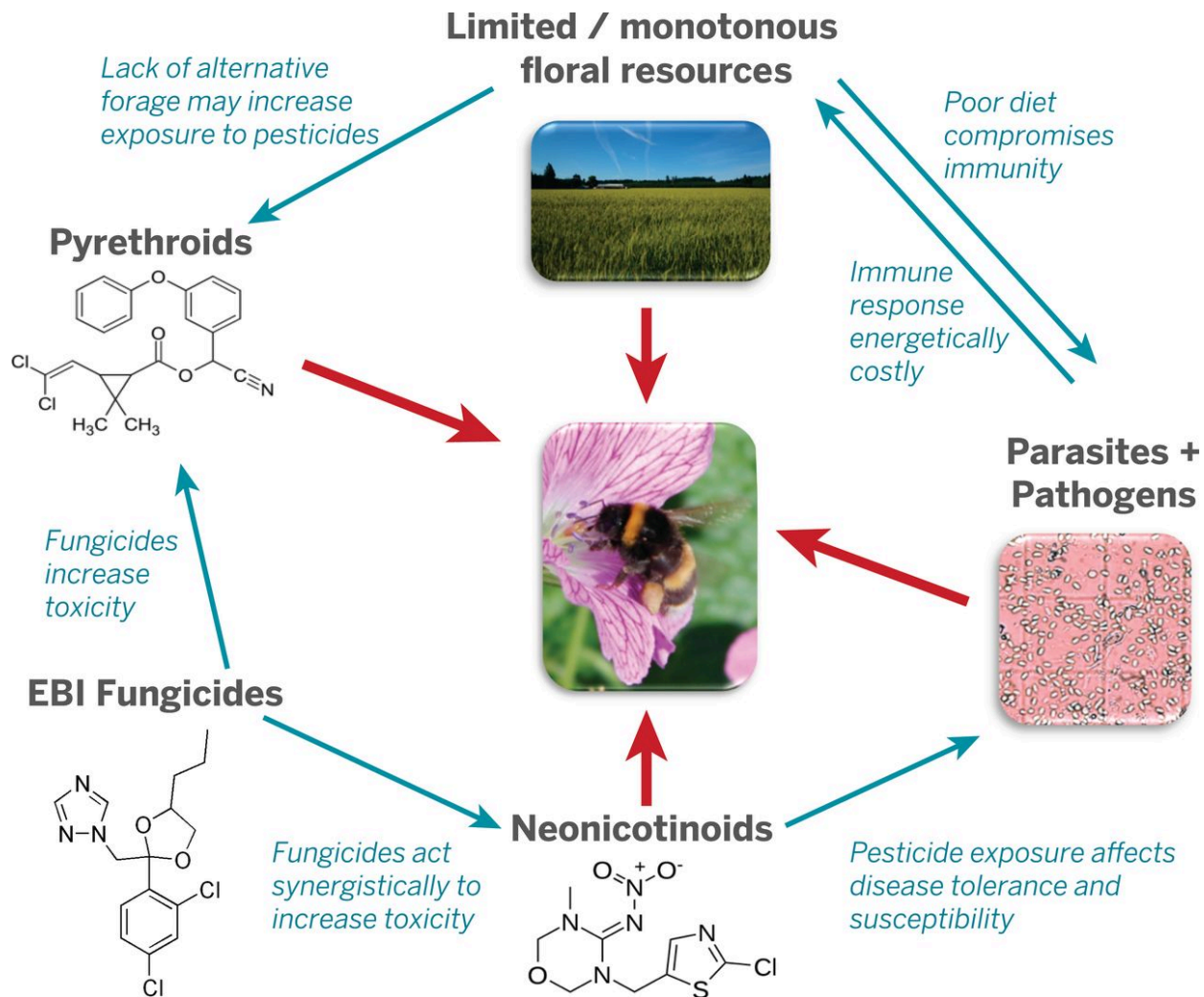
Specific immune responses in mice following subchronic exposure to acetamiprid

Soumaya Marzouki^{a,1}, Ines Bini Dhouib^{b,c,1}, Chaouki Benabdessalem^a, Raja Rekik^a, Raoudha Doghri^d, Ammar Maroueni^e, Zakaria Bellasfar^e, Saloua Fazaa^c, Jihene Bettaieb^{a,f}, Mohamed Ridha Barbouche^{a,f}, Melika Ben Ahmed^{a,f,*}

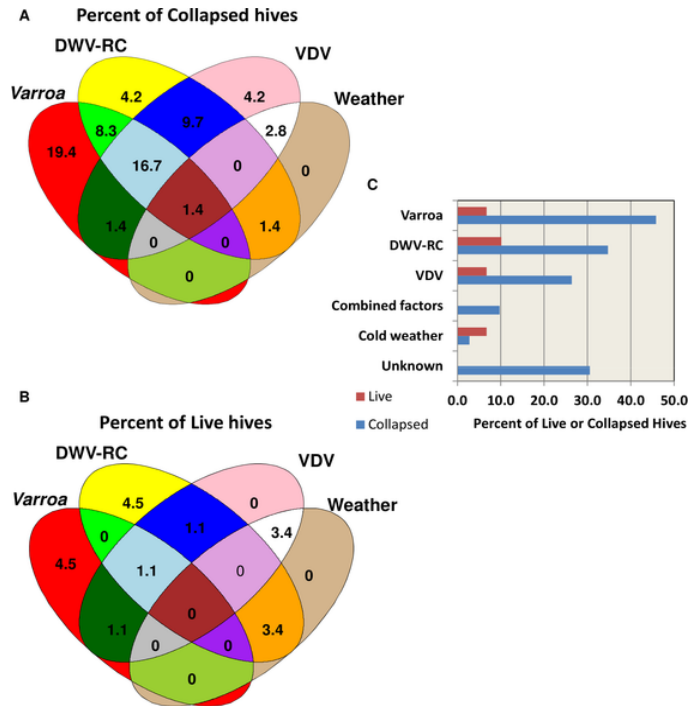
Results: The ACE-treated mice showed a significant immunosuppression of the specific humoral response with a restorative effect of curcumin when administered with ACE. Similarly, ACE significantly decreased the level of splenocyte proliferation after either a non specific or a specific activation. Curcumin partially restores the antigen specific cellular immune response. Moreover, when administered alone, curcumin significantly inhibits the proliferative responses to the mitogen confirming its anti-mitogenic effect. Histological analysis showed alteration of spleens of mice exposed to ACE.

Significance: Altogether, our data indicated that ACE could potentially be harmful to the immune system.

Wild and managed bees are exposed to a number of interacting stressors



Multifactorial induction of hive collapse

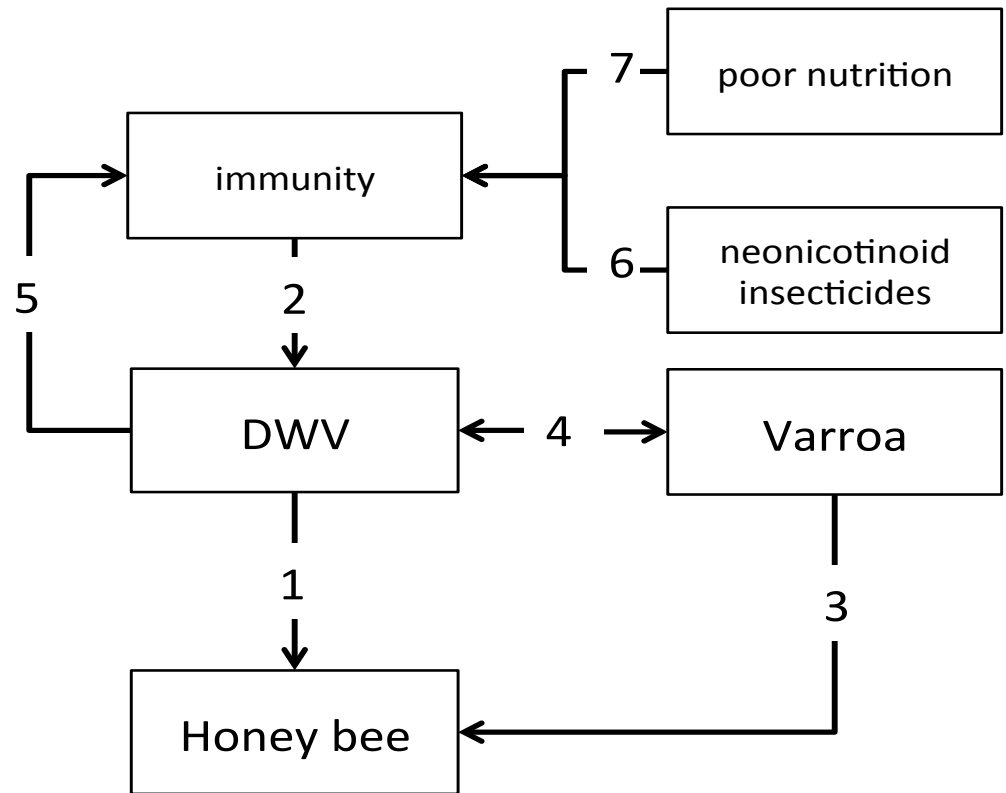


- ✓ Varroa infestation and DWV replication contribute to 70% of colony losses
- ✓ A single factor may not be sufficient to trigger colony losses
- ✓ A combination of stressors appears to impact hive health

Conclusions

- ✓ DWV mediates honeybee immunosuppression by targeting NF- κ B signaling
- ✓ *Varroa* mites promote DWV replication, exacerbate immunosuppression and enhance their fitness
- ✓ Neonicotinoids upregulate an inhibitor of NF- κ B activation and triggers immunosuppression, which promotes DWV replication
- ✓ Nutrition cross-modulates honey bee immune pathways

The “Sword of Damocles” paradigm





University of Napoli “Federico II”
Department of Agricultural Sciences



Thank you!